

# Lipase mediated preparation of the enantiomers of 3,3,3-trifluoro-2-methylpropanoic acid

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**Abstract**—The preparation of both enantiomers of 3,3,3-trifluoro-2-methylpropanoic acid by a lipase mediated kinetic resolution of the racemate is described.

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## 1. Introduction

Compounds containing the trifluoromethyl group have and continue to play an important role as fine chemical constituents contributing to the development of pharmaceutical, agrochemical and materials products.<sup>1</sup> Many of these are nonchiral aryl CF<sub>3</sub> and OCF<sub>3</sub> containing compounds, however there has been the recent emergence of commercial products carrying the CF<sub>3</sub> group at a stereogenic centre.<sup>2</sup> The number of available starting materials in this category is few and methods for preparing compounds with the CF<sub>3</sub> at a stereogenic centre are limited.<sup>3</sup> Herein we report the lipase mediated resolution of 3,3,3-trifluoro-2-methylpropanoic acid **1**. The only reported preparation of enantioenriched **1** was achieved by hydrogenation of 2-(trifluoromethyl)acrylic acid with H<sub>2</sub> in the presence of Ru-(*R*)-BINAP catalyst. However product **1** was not isolated and characterized, but directly transformed into (2*S*)-3,3,3-trifluoro-2-methyl-1-propanol (ee 80%).<sup>4</sup> Some  $\alpha$ -trifluoromethylated carboxylic acids have been obtained in optically active form (unknown stereochemistry) by enzymatic hydrolysis of their esters using the PS lipase in an aqueous medium,<sup>5</sup> yet the resolution of **1** has not been described. We have been exploring the resolution of fluorinated esters in dry organic solvents<sup>6</sup> and in this program decided to address the resolution of **1** by

esterification using the *Candida rugosa* lipase<sup>†</sup>(CRL) in hexane. This was an attractive method as it allowed us to test the ability of such enzymes to distinguish directly between the CH<sub>3</sub> and CF<sub>3</sub> groups. Previous analyses and studies in asymmetric syntheses suggest a significant difference in the steric influence of a CH<sub>3</sub> and CF<sub>3</sub> group,<sup>7</sup> with an estimate that the CF<sub>3</sub> approximates the size of an isopropyl group.<sup>8</sup> It is somewhat surprising that this has not been explored in this system before. Herein we have successfully resolved each of the enantiomers in high enantiomeric excess.

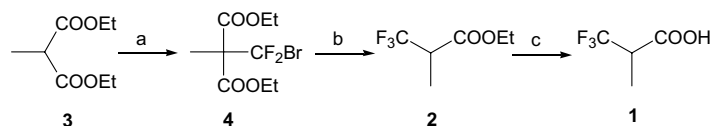
## 2. Results and discussion

Ethyl 3,3,3-trifluoro-2-methylpropanoate **2** was prepared in two steps by the reaction of diethyl 2-methylmalonate **3** with sodium hydride and dibromo(difluoro)methane followed by a fluorination–decarboxylation process.<sup>9</sup> Alkaline hydrolysis of the resultant ester proved unsuccessful leading to conversion of the CF<sub>3</sub> group to a carbonylate with the generation of 2-methylmalonate.<sup>10</sup> However acidic hydrolysis gave the desired 3,3,3-trifluoro-2-methylpropanoic acid,<sup>11</sup> **1** as illustrated in Scheme 1.

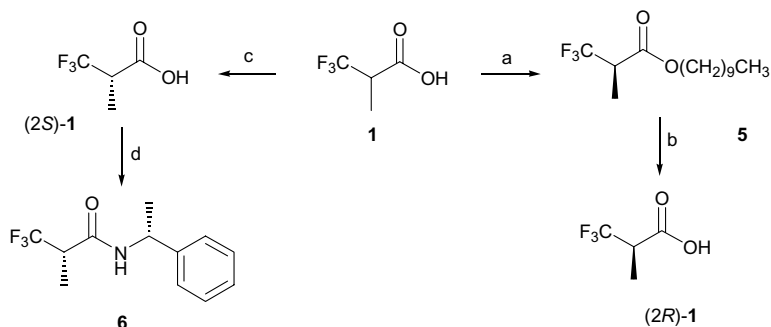
Lipase mediated kinetic resolution of **1** via enantioselective esterification with 1-decanol in hexane gave decyl ester **5**. When this reaction was stopped after 4h

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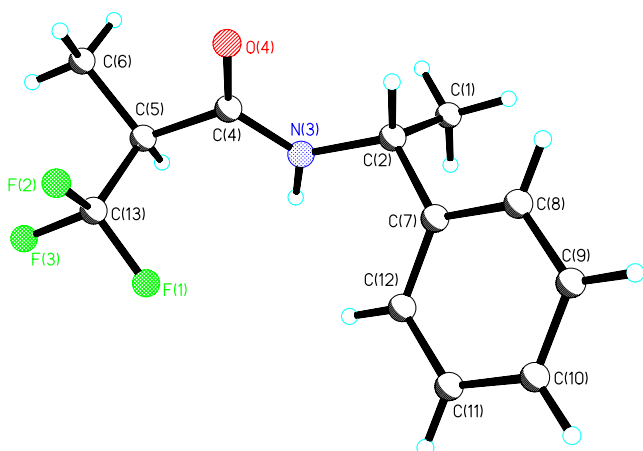
<sup>†</sup>Lipase from *Candida rugosa*, Type VII (CRL) was purchased from the Sigma Chemical Co. and had a specific activity of 1140 U mg<sup>-1</sup> solid. The lipase was used 'straight from the bottle'.



**Scheme 1.** Reagents and conditions: (a) NaH,  $\text{CF}_2\text{Br}_2$ , THF, 48%; (b) KF, DMSO, 170°C, 91%; (c) HCl, 1,4-dioxane, reflux, 61%.



**Scheme 2.** Reagents and conditions: (a) CRL, 1-decanol (2equiv), hexane, 37°C, 4h; (b) HCl, 1,4-dioxane, reflux; (c) CRL, 1-decanol (2equiv), hexane, 37°C, 20h; (d) *N*-methylmorpholine, ClCOOMe, (1*R*)-1-phenylethylamine, THF, −10°C, 90%.



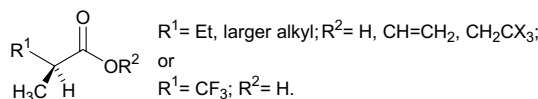
**Figure 1.** X-ray derived structure of (2*S*)-3,3,3-trifluoro-2-methyl-*N*-[(1*R*)-1-phenyl]propanamide **6**.<sup>‡</sup>

and **5** was hydrolyzed, enantiomerically enriched **1** was obtained<sup>12</sup> ee 90%, 36%,  $[\alpha]_{\text{D}}^{19} < +1$  ( $c$  1.12, MeOH) as shown in Scheme 2. Longer reaction times (20 h) gave the other enantiomer of **1** recovered as the unreacted and enantiomerically pure acid<sup>13</sup> {ee >98%, 74%,  $[\alpha]_{\text{D}}^{19} = -0.8$  ( $c$  1.23, MeOH)}, Scheme 2.<sup>§</sup>

To determine the absolute configuration of the recovered **1**, amide **6**  $\{[\alpha]_{\text{D}}^{25} = +88.7$  ( $c$  0.345,  $\text{CHCl}_3$ ) $\}$  was prepared<sup>14,15</sup> by reaction with (1*R*)-1-phenylethylamine, (Scheme 2). X-ray diffraction analysis of a single crystal of amide **6** showed an (*S*)-configuration on the acid de-

rived moiety (Fig. 1) and therefore a lipase preference for enantiomer (2*R*)-**1**.

The stereochemical outcome of the CRL catalyzed kinetic resolution follows the pattern of reactivity of 2-methylalkanoic acids.<sup>16</sup> The faster reacting enantiomers have a stereochemistry as shown in Scheme 3, a configurational preference that is persistent through enzymatic hydrolysis, esterification or transesterification reactions with this lipase.



**Scheme 3.** The stereochemistry of the faster reacting enantiomer with CRL.

## References and notes

- Chambers, R. D. *Fluorine in Organic Chemistry*; Blackwell: Abingdon, UK, 2004.
- (a) Thompson, A. S.; Corley, E. G.; Huntington, M. F.; Grabowski, E. J. J. *Tetrahedron Lett.* **1995**, *36*, 8937; (b) Thompson, A. S.; Corley, E. G.; Huntington, M. F.; Grabowski, E. J. J.; Remenar, J. F.; Collum, D. B. *J. Am. Chem. Soc.* **1998**, *120*, 2028; (c) Pierce, M. E.; Parsons, R. L.; Radesca, L. A.; Lo, Y. S.; Silverman, S.; Moore, J. R.; Islam, Q.; Choudhury, A.; Fortunak, J. M. D.; Nguyen, D.; Luo, C.; Morgan, S. J.; Davis, W. P.; Canfalone, P. N.; Chen, C.; Tillyer, R. D.; Frey, L.; Tan, L.; Xu, F.; Zhao, D.; Thompson, A. S.; Corley, E. G.; Grabowski, E. J. J.; Reamer, R.; Reider, P. J. *J. Org. Chem.* **1998**, *63*, 8536.
- Enantiocontrolled Synthesis of Fluoro-organic Compounds*; Soloshonok, V. A., Ed.; Wiley: Chichester, 1999.
- (a) Iseki, K.; Kuroki, Y. T.; Nagai, T.; Kobayashi, Y. *Chem. Pharm. Bull.* **1996**, *44*, 477; (b) Iseki, K.; Kuroki, Y.; Nagai, Y.; Kobayashi, Y. *J. Fluorine Chem.* **1994**, *69*, 5.
- Watanabe, S. *J. Fluorine Chem.* **1992**, *59*, 249.
- Beier, P.; O'Hagan, D. *J. Chem. Soc., Chem. Commun.* **2002**, 1680.

<sup>‡</sup> The crystallographic data (excluding structure factors) for **6** have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 236631.

<sup>§</sup> The ee values of the enantiomers of **1** were determined by GC–MS using a chiral β-DEX 120 (Supelco) column (30 m × 250 μm, film thickness 0.25 μm). Temp 75°C hold 20 min, then at 10°C min<sup>−1</sup> to 100°C, hold 11 min, ret. time 29.5 min (*R*), 30.15 min (*S*).

7. O'Hagan, D.; Rzepa, H. S. *J. Chem. Soc., Chem. Commun.* **1997**, 645.
8. (a) Seebach, D. *Angew. Chem. Int., Ed. Engl.* **1990**, *29*, 1320; (b) Bott, G.; Field, L. D.; Sternhill, S. *J. Am. Chem. Soc.* **1980**, *102*, 5618.
9. Everet, T. S.; Purrington, S. T.; Bumgardner, C. L. *J. Org. Chem.* **1984**, *49*, 3702.
10. Buxton, M. W.; Stacey, M.; Tatlow, J. C. *J. Chem. Soc.*, **1954**, 366.
11. 3,3,3-Trifluoro-2-methylpropanoic acid, **1**: **2** (16.1 g, 9.7 mmol) was dissolved in 1,4-dioxane (100 cm<sup>3</sup>) and HCl (36%, 70 cm<sup>3</sup>) added. The mixture was heated under reflux for 15 h, cooled and DCM (200 cm<sup>3</sup>) then added. The two phases were separated and the aqueous was extracted into DCM (3 × 100 cm<sup>3</sup>). The combined organic extracts were washed with aq NaHCO<sub>3</sub> (0.8 M, 3 × 100 cm<sup>3</sup>). The aqueous phase was acidified (conc. HCl), the product isolated by extraction into diethyl ether (3 × 100 cm<sup>3</sup>), drying over MgSO<sub>4</sub> and removal of the solvent under reduced pressure. Distillation (80 °C/20 mm Hg) gave **1** as a colourless liquid (8.205 g, 61%). MS (CI): *m/z* = 143.0316 [M+H]<sup>+</sup> C<sub>4</sub>H<sub>6</sub>O<sub>2</sub>F<sub>3</sub> requires 143.0320. IR (film) 2923, 2853, 2689, 1701 (CO), 1471, 1327, 1083, 1043, 965, 702 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>; Me<sub>4</sub>Si): δ 9.9 (br s, 1H), 3.33–3.18 (m, 1H), 1.47 (d, 3H, *J* = 7.2 Hz). <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>; CFCl<sub>3</sub>): δ -70.5 (d, *J* = 6.9 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 173.3, 124.7 (q, *J* = 279 Hz), 44.4 (q, *J* = 29 Hz), 11.0. MS (EI): *m/z* (rel int.) = 142 [M]<sup>+</sup> (1), 125 (30), 122 [M-F]<sup>+</sup> (20), 102 (35), 78 (70), 77 (100), 69 [CF<sub>3</sub>]<sup>+</sup> (20), 45 (35).
12. (2*R*)-3,3,3-Trifluoro-2-methylpropanoic acid, (2*R*)-**1**: 1-Decanol (6.708 g, 42.38 mmol) and **1** (3.009 g, 21.19 mmol) were dissolved in hexane (200 cm<sup>3</sup>). The CRL (7.5 g) was added and the mixture shaken at 250 rpm/37 °C for 4 h. The reaction was terminated by the addition of MgSO<sub>4</sub> (5 g) and the mixture filtered. The solid was washed with diethyl ether (3 × 15 cm<sup>3</sup>), the solvent removed under reduced pressure and the residue dissolved in diethyl ether (50 cm<sup>3</sup>). The solution was washed with aq NaHCO<sub>3</sub> (0.8 M, 3 × 40 cm<sup>3</sup>), the solvent removed from the organic phase and the residue hydrolyzed (HCl in 1,4-dioxane). Similar work-up as for the racemic acid afforded (2*R*)-**1** (541 mg, 36%, ee 90%), [α]<sub>D</sub><sup>19</sup> < +1 (*c* 1.12, MeOH).
13. (2*S*)-3,3,3-Trifluoro-2-methylpropanoic acid, (2*S*)-**1**: 1-Decanol (6.708 g, 42.38 mmol) and **1** (3.009 g, 21.19 mmol) were dissolved in hexane (200 cm<sup>3</sup>). The CRL (7.5 g) was added and the mixture shaken at 250 rpm/37 °C for 20 h. The reaction was terminated by the addition of MgSO<sub>4</sub> (5 g) and the mixture filtered. The solid was washed with diethyl ether (3 × 15 cm<sup>3</sup>), the solvent removed under reduced pressure and the residue dissolved in diethyl ether (50 cm<sup>3</sup>). The unreacted acid was isolated by extraction with aq NaHCO<sub>3</sub> (0.8 M, 3 × 40 cm<sup>3</sup>), acidification and extraction into diethyl ether (3 × 20 cm<sup>3</sup>). Drying and solvent removal gave (2*S*)-**1** (1.111 g, 74%, ee >98%). [α]<sub>D</sub><sup>19</sup> = -0.8 (*c* 1.23, MeOH).
14. Compagnone, R. S.; Rapoport, H. *J. Org. Chem.* **1986**, *51*, 1713.
15. (2*S*)-3,3,3-Trifluoro-2-methyl-*N*-[(1*R*)-1-phenyl]-propanamide, **6**: (2*S*)-**1** (ee >98%, 106 mg, 0.746 mmol) was dissolved in dry THF (12 cm<sup>3</sup>) and cooled to -10 °C under nitrogen. 4-Methylmorpholine (81 mg, 0.8 mmol) and methyl chloridocarbonate (75 mg, 0.8 mmol) were added and after 1 min of stirring (1*R*)-1-phenylethanamine (113 mg, 0.93 mmol) then added. The solution was stirred at -5 to -10 °C for 40 min and then an aqueous solution of citric acid (5%, 150 cm<sup>3</sup>) added. The mixture was extracted with ethyl acetate (3 × 40 cm<sup>3</sup>), the organic phase washed with aqueous solution of sodium hydrogen carbonate (1 M, 3 × 20 cm<sup>3</sup>), dried over MgSO<sub>4</sub> and the solvent removed under reduced pressure. The residue (de 98%) was purified using silica gel column chromatography to give the major diastereomer as a white solid (de 100%, 165 mg, 90%) mp 138.5–139 °C (from hexane). [α]<sub>D</sub><sup>25</sup> = +88.7 (*c* 0.345, CHCl<sub>3</sub>). MS (CI): *m/z* = 246.1107 [M+H]<sup>+</sup> C<sub>12</sub>H<sub>15</sub>OF<sub>3</sub>N requires 246.1107. IR (KBr) 3308, 3093, 2987, 2972, 1651 (CO), 1560, 1268, 1239, 1175, 1125, 1006, 754, 701 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>; Me<sub>4</sub>Si): δ 7.38–7.25 (m, 5H), 5.9 (br s, 1H), 5.19–5.09 (m, 1H), 3.09–2.93 (m, 1H), 1.51 (d, 3H, *J* = 6.8 Hz), 1.39 (d, 3H, *J* = 7.2 Hz). <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>; CFCl<sub>3</sub>): δ -69.9 (d, *J* = 8.6 Hz). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 165.6, 142.4, 128.8, 127.6, 126.0, 125.6 (q, *J* = 280 Hz), 49.2, 46.3 (q, *J* = 28 Hz), 21.5, 11.0. MS (EI): *m/z* (rel int.) = 245 [M]<sup>+</sup> (60), 230 [M-Me]<sup>+</sup> (50), 120 (20), 106 (100), 105 (40), 77 (30). MS (ESI): *m/z* = 244 [M-H]<sup>-</sup>.
16. (a) Engel, K. H. *Tetrahedron: Asymmetry* **1991**, *2*, 165; (b) Berglund, P.; Holmquist, M.; Hedenström, E.; Hult, K.; Högberg, H. E. *Tetrahedron: Asymmetry* **1993**, *4*, 1869.